# Synthesis, Resolution, and Preliminary Evaluation of trans-2-Amino-6(5)-hydroxy-1-phenyl-2,3-dihydro-1H-indenes and Related **Derivatives as Dopamine Receptors Ligands**

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The present work reports the synthesis of enantiomeric pairs of the *trans*-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene [(+)-**14a**, (-)-**14a**] and *trans*-2-amino-5-hydroxy-1-phenyl-2,3dihydro-1*H*-indene [(+)-**14b**, (-)-**14b**] and their *N*,*N*-di-*n*-propyl [(+)- and (-)-**15a,b**], *N*-methyl-N-allyl [(+)- and (-)-16a,b], and N-methyl-N-n-propyl [(+) and (-)-17a,b] derivatives obtained by a combination of stereospecific reactions and optical resolution. The new compounds were evaluated for their affinity at the dopamine  $D_1$  and  $D_2$  receptors. The amines (+)- and (-)-14a, incorporating the D<sub>1</sub> pharmacophore 2-phenyl-2-(3-hydroxyphenyl)ethylamine in a *trans* extended conformation, and their derivatives displayed  $D_1$  and  $D_2$  affinity in the nanomolar range. On the other hand, the enantiomers (+)- and (-)-**14b**, (+)- and (-)-**15b** displayed high affinity and selectivity for the  $D_1$  receptor. In a preliminary behavioral study on rats (+)-14b, and to a greater extent (+)-15b, promoted episodes of intense grooming, thus indicating that they act as central  $D_1$  agonists. The *trans*-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indenes (+)-14b and (+)-15b represent selective D<sub>1</sub> agonists lacking a catechol group, which should meet the prerequisites for a central nervous system penetration.

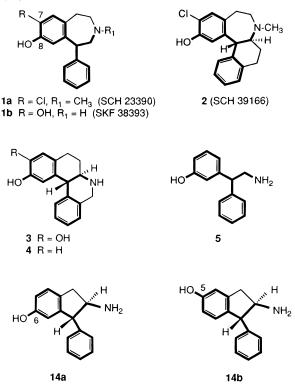
# Introduction

The physiological effects of the neurotransmitter dopamine (DA) are mediated by receptors, the dysfunction of which is implicated in psychiatric, neurological, and neuroendocrine disorders such as schizophrenia, Parkinson's disease, and hyperprolactinemia. At present, the classification commonly accepted divides DA receptors into two families:  $D_{1-like}$  and  $D_{2-like}$ . The  $D_{1-like}$ family includes the D<sub>1</sub> and D<sub>5</sub> receptors, whereas the  $D_2$ ,  $D_3$ , and  $D_4$  receptors belong to the  $D_{2-like}$  family.<sup>1</sup>

While many researches were addressed on the discovery of new selective therapeutic agents acting on  $D_{2-like}$  receptors, the development of selective agonists or antagonists for D<sub>1-like</sub> receptors has received less attention. The discovery by Schering-Plough Research of the (R)-(+)-7-chloro-8-hydroxy-3-methyl-1-phenyl-2,3,4,5-tetrahydro-1H-3-benzazepine (1a, SCH 23390, Chart 1) and the introduction of its tritiated derivative as radioligand have facilitated research on the function of  $D_{1-like}$  receptors in the central nervous system. Compound **1a** is the first selective  $D_{1-like}$  receptor antagonist and the prototype of the 1-phenyl-3-benzazepine class which includes both agonists and antagonists.<sup>2</sup> The specific D<sub>1-like</sub> antagonist, (-)-trans-11chloro-6,6a,7,8,9,13b-hexahydro-7-methyl-5H-benzo-[d]naphtho[2,1-b]azepin-12-ol (2, SCH 39166)<sup>3</sup> has been proposed subsequently as a potential antipsychotic drug for the treatment of schizophrenia without acute extrapyramidal side effects (EPS).<sup>4</sup>

Dopaminergic 1-phenyl-3-benzazepines display a high degree of enantioselectivity, and the activity resides only





in an enantiomer as observed also in 2. This compound can exist as cis and trans isomers and contains two chiral carbons. In binding studies, the trans isomer showed  $D_{1-like}$  affinity higher than that of the *cis* isomer. Among the enantiomers of 2, the compound with a 6aS,-13bR configuration showed high affinity for the D<sub>1-like</sub> receptors. A similar situation was observed in the hexahydrobenzo[a]phenanthridine class in which only the trans isomer 3 with a 6aR,12bS configuration has

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## 2,3-Dihydro-1H-indenes as Dopamine Receptor Ligands

high affinity for  $D_{1-like}$  receptors.<sup>5</sup> In both classes, the presence of a catechol function induces agonistic activity (**1b**<sup>6</sup> and **3**), while a single hydroxyl group at a position meta to the ethylamine chain is a prerequisite for  $D_{1-like}$  binding and antagonistic activity (**1a**, **2**, and **4**).<sup>7</sup>

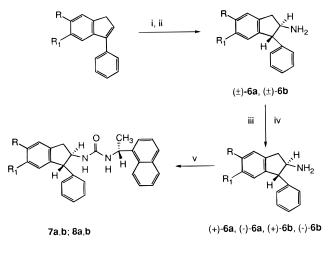
Nichols and Brewster have proposed a conformational model for the  $D_1$  receptor, suggesting that the hydroxyphenylethylamine moiety embedded in the hexahydrobenzo[*a*]phenanthridines **3** and **4** should be in a *trans* extended conformation.<sup>5</sup> This structural feature is probably an important contributor to the  $D_{1-like}$  selectivity. The unsubstituted phenyl ring seems to be responsible for the induction of  $D_{1-like}$  selectivity as well as for an increased affinity and potency by interacting with an accessory binding region of the receptor.<sup>8</sup> On the basis of the structure of cloned catecholaminergic G protein receptors,<sup>1</sup> the same receptor model postulates that an aspartate residue binds the protonated amino group and that the hydroxyls of two serine residues bind the catechol function.<sup>8,9</sup>

On the basis of these findings, we have synthesized the enantiomers of *trans* -2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene ((+)- and (-)-**14a**, Chart 1), which contain the putative pharmacophore of  $D_{1-like}$  receptor, the 2-phenyl-2-(3-hydroxyphenyl)ethylamine **5** with a *trans* conformation.<sup>5</sup> Compound **14a** contains an amino group *trans* to the 1-phenyl ring and two asymmetric centers (C1 and C2) and produces two enantiomers.

The structure–activity relationships (SAR) on 1-phenyl-3-benzazepines indicate that the 8-hydroxy group is a critical pharmacophoric element, but the conceptual model of the  $D_{1-like}$  receptor proposes that also the 7-hydroxy group interacts with a serine residue and is determinant for the  $D_{1-like}$  agonistic activity.<sup>8</sup> In view of this, we also synthesized the enantiomers of *trans*-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene ((+)and (-)-**14b**). In both compounds, **14a** and **14b**, the amino group was substituted with methyl, allyl, and n-propyl groups which, as a rule, affect the affinity for DA receptors.<sup>10a,b</sup>

## Chemistry

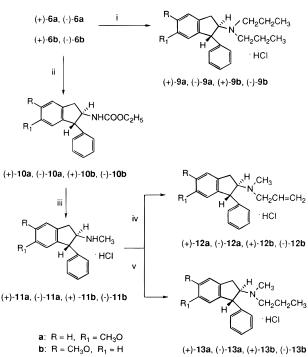
The new compounds were synthesized from 5- or 6-methoxy-3-phenyl-1H-indene prepared by reaction of 6- or 5-methoxyindan-1-one with phenylmagnesium bromide according to the procedure reported in the literature for 3-phenyl-1*H*-indene<sup>11</sup> (Scheme 1). The methoxy-3-phenyl-1H-indenes were converted to trans-2-amino-6(5)-methoxy-1-phenyl-2,3-dihydro-1H-indenes  $(\pm)$ -**6a** and  $(\pm)$ -**6b** by a stereospecific hydroboration and reaction of organoboranes with hydroxylamine *O*-sulfonic acid.<sup>12</sup> The *trans* configuration of  $(\pm)$ -**6a** and  $(\pm)$ -**6b** was established on the *cis* addition mechanism of the diborane and the <sup>1</sup>H-NMR data. In the NMR spectra of the two compounds, we noted a coupling constant of 7.93 Hz for the hydrogens at the 1- and 2-positions. This value is very similar to that reported in literature (8 Hz) for the same hydrogens in trans-2amino-1-phenyl-2,3-dihydro-1H-indene.13 The resolution of the enantiomers of  $(\pm)$ -**6a** and  $(\pm)$ -**6b** was accomplished by recrystallization of the (+)- and (-)-Ldibenzoyltartaric acid salts. The free bases were obtained by treatment of the salts with aqueous ammonia. The enantiomeric excess was determined derivatizing Scheme 1<sup>a</sup>



**a**: R = H,  $R_1 = CH_3O$ **b**:  $R = CH_3O$ ,  $R_1 = H$ 

 $^a$  (i) NaBH4, BF3; (ii) NH2OSO3H; (iii) (–)-dibenzoyl-L-tartaric acid; (iv) (+)-dibenzoyl-L-tartaric acid; (v) (R)-(–)-1-(1-naphthyl)-ethyl isocyanate.

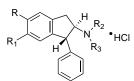
Scheme 2<sup>a</sup>



 $^{a}$  (i) Propionic acid, NaBH<sub>4</sub>, HCl; (ii) ethyl chloroformate, triethylamine; (iii) LiAlH<sub>4</sub>, HCl; (iv) allyl bromide, K<sub>2</sub>CO<sub>3</sub>, HCl; (v) *n*-propyl iodide, K<sub>2</sub>CO<sub>3</sub>, HCl.

the amines with (*R*)-1-(1-naphthyl)ethyl isocyanate.<sup>14</sup> The inspection and integration of the benzylic and methyl protons signals in the diastereomeric ureas **7** and **8** provided an enantiomeric excess greater than 98%. The di-*n*-propyl derivatives **9** were obtained by alkylation of the enantiomeric free bases **6a,b** with the NaBH<sub>4</sub>-propionic acid complex.<sup>15</sup> The *N*-*n*-propyl-*N*-methyl and *N*-allyl-*N*-methyl derivatives were prepared as outlined in Scheme 2. Cleavage of the methoxyl groups was achieved by employing the gentle methion-ine-methanesulfonic acid procedure.<sup>16</sup> The hydroxy derivatives **14a,b-17a,b** are listed in Table 1.

**Table 1.** Affinities of *trans*-2-Amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indenes and *trans*-2-Amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indenes at Striatal Dopamine Receptors<sup>a</sup>



| compd                      | R  | $R_1$ | R <sub>2</sub>  | R <sub>3</sub>                    | K <sub>i</sub> (nM) |                 |
|----------------------------|----|-------|-----------------|-----------------------------------|---------------------|-----------------|
|                            |    |       |                 |                                   | D1                  | $D_2$           |
| (+)- <b>14a</b>            | Н  | OH    | Н               | Н                                 | $2.5\pm0.2$         | $0.59\pm0.06$   |
| (–)- <b>14a</b>            | Н  | OH    | Н               | Н                                 | $34.5\pm2.4$        | $0.89\pm0.07$   |
| (+)- <b>15a</b>            | Н  | OH    | $(CH_2)_2CH_3$  | $(CH_2)_2CH_3$                    | $28.5\pm1.6$        | $51.5\pm2.3$    |
| (–)-15a                    | Н  | OH    | $(CH_2)_2CH_3$  | $(CH_2)_2CH_3$                    | $26.4\pm2.5$        | $0.84\pm0.06$   |
| (+)- <b>16a</b>            | Н  | OH    | CH <sub>3</sub> | CH <sub>2</sub> CHCH <sub>2</sub> | $3.0\pm0.3$         | $37.5\pm1.4$    |
| (–)-16a                    | Н  | OH    | CH <sub>3</sub> | CH <sub>2</sub> CHCH <sub>2</sub> | $0.79\pm0.09$       | $0.43\pm0.03$   |
| (+)- <b>17a</b>            | Н  | OH    | CH <sub>3</sub> | $(CH_2)_2CH_3$                    | $0.90\pm0.08$       | $0.30\pm0.03$   |
| (–)-17a                    | Н  | OH    | $CH_3$          | $(CH_2)_2CH_3$                    | $0.88\pm0.09$       | $0.34\pm0.02$   |
| (+)- <b>14b</b>            | OH | Н     | Н               | H                                 | $1.87\pm0.06$       | >1000           |
| (–)-14b                    | OH | Н     | Н               | Н                                 | $18.7\pm0.8$        | >1000           |
| (+)- <b>15b</b>            | OH | Н     | $(CH_2)_2CH_3$  | $(CH_2)_2CH_3$                    | $1.4\pm0.1$         | >1000           |
| (–)- <b>15b</b>            | OH | Н     | $(CH_2)_2CH_3$  | $(CH_2)_2CH_3$                    | $1.76\pm0.07$       | >1000           |
| (+)- <b>16b</b>            | OH | Н     | CH <sub>3</sub> | CH <sub>2</sub> CHCH <sub>2</sub> | >1000               | >1000           |
| (–)- <b>16b</b>            | OH | Н     | $CH_3$          | CH <sub>2</sub> CHCH <sub>2</sub> | >1000               | >1000           |
| (+)- <b>17b</b>            | OH | H     | CH <sub>3</sub> | $(CH_2)_2CH_3$                    | >1000               | >1000           |
| (–)- <b>17b</b>            | OH | Н     | $CH_3$          | $(CH_2)_2CH_3$                    | >1000               | >1000           |
| 18                         | OH | OH    | Н               | H                                 | $5.5\pm0.1$         | >1000           |
| DA                         |    |       |                 |                                   | >1000               | >1000           |
| ( <i>R</i> )-(+)-SCH 23390 |    |       |                 |                                   | $0.73\pm0.05$       | >1000           |
| PPHT                       |    |       |                 |                                   | >1000               | $2.70 \pm 0.12$ |

<sup>*a*</sup> Binding studies were performed using frozen sections of rat neostriatum with [<sup>3</sup>H]SCH 23390 (D<sub>1</sub> selective antagonist) and [<sup>3</sup>H]spiperone (D<sub>2</sub> selective antagonist) as radioligands.  $K_i$  represents the inhibition constant which was calculated according to Cheng and Prusoff. Values are the means  $\pm$  SEM of five to seven independent triplicate experiments.

## **Biological Results and Discussion**

The target compounds were screened for their *in vitro* binding affinity to DA receptors, using sections of rat neostriatum according to the technique previously described.<sup>17</sup> Rat neostriatum expresses primarily D<sub>1</sub> and D<sub>2</sub> subtypes.<sup>18</sup> The radioligands [<sup>3</sup>H]SCH 23390 (D<sub>1</sub> selective) and [<sup>3</sup>H]spiperone (D<sub>2</sub> selective) were used, while the reference compounds used were (*R*)-(+)-SCH 23390 (a selective D<sub>1</sub> antagonist), (±)-2-(*N*-(phenyl-ethyl)-*N*-propylamino)-5-hydroxytetralin hydrochloride (PPHT, a selective D<sub>2</sub> agonist), and DA (Table 1).

Data obtained for 2-amino-6-hydroxy-1-phenyl-1Hindenes (+)- and (-)-14a-17a, which hold the pharmacophore 2-phenyl-2-(3-hydroxyphenyl)ethylamine in a trans conformation, indicate that these compounds interact with both DA receptor subtypes. The primary amines (+)-14a, (-)-14a show higher affinity for the D<sub>2</sub> than for the  $D_1$  receptor, and (-)-14a is 38 times more selective for the D<sub>2</sub> receptor. Introduction of two alkyl substituents on the amino group influences in a different way the affinity for  $D_1$  and  $D_2$  receptors. Dipropylation of (+)-14a decreases  $D_1$  and  $D_2$  affinity, while the same substitution on (-)-14a does not affect  $D_1$  and  $D_2$ affinity. The introduction of a methyl and an allyl group on (+)-14a decreases only  $D_2$  affinity, and (+)-16a is selective for  $D_1$  receptor (12 times), while the same substitution on (-)-14a increases only  $D_1$  affinity. Methyl and propyl introduction on the amino group increases D<sub>1</sub> affinity, providing compounds [(+)-17a and (-)-17a] with high affinity for both receptors. Thus, the reduction of alkyl group dimensions (from propyl to methyl) increases D<sub>1</sub> affinity. Moreover, different affinities were observed for (+)- and (-)-**14a** at D<sub>1</sub> receptor, for (+)- and (-)-15a at D<sub>2</sub> receptor, and for (+)and (-)-**16a** at D<sub>1</sub> and D<sub>2</sub> receptors.

These results indicate that the compounds (+)- and (-)-**14a**-**17a** are not highly selective, displaying D<sub>1</sub> and D<sub>2</sub> affinity. This suggests that the 2-phenyl-2-(3-hydroxyphenyl)ethylamine pharmacophore embedded in these compounds in a trans conformation does not induce high  $D_1$  selectivity. This finding could be explained on the basis of the structural differences among our compounds, having an exocyclic amino group, and tetrahydrobenzazepines or hexahydrophenanthridines, which hold the pharmacophore in a more rigid framework. Conformational analysis and SAR in the benzazepines series suggest that the biologically active conformation of the tetrahydrobenzazepine ring is a chair conformation with an equatorial phenyl ring.<sup>19</sup> In contrast, an axial phenyl ring is detrimental to  $D_1$ receptor affinity.<sup>3</sup> In an interesting study on the SAR of the benzo[*a*]phenanthridines **3** and **4**, Brewster et al. concluded that the unsubstituted phenyl ring attached at the 2-position of the ethylamine side chain is nearly coplanar with the catechol ring.<sup>20</sup>

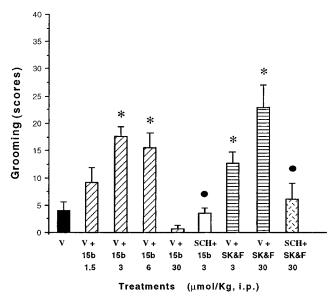
In the *trans*-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indenes (+)- and (-)-**14a**-**17a**, the unsubstituted phenyl ring and the amino group are not rigidly constrained, and a free rotation is allowed. Thus, the orientation of the two groups can deviate from that preferred by the D<sub>1</sub> receptor and the two phenyl ring may be not coplanar. A similar situation could be responsible for the moderate affinity observed in *trans*-1-phenyl-2-amino-6,7-dihydroxytetralin for both D<sub>1</sub> and D<sub>2</sub> receptors.<sup>20</sup> Moreover, on the basis of current models, the D<sub>1</sub> receptor requires the catechol function for high affinity, while the D<sub>2</sub> receptor is much less demanding, and only a single hydroxy group is necessary to confer affinity.<sup>5,20-22</sup> Thus, these considerations could explain the low selectivity of the monophenolic compounds (+)- and (–)-**14a**–**17a** for  $D_1$  and  $D_2$  receptors.

An unexpected result was obtained with the trans-2amino-5-hydroxy-1-phenyl-2,3-dihydro-1H-indenes. In this series, the primary amines (+)-14b, (-)-14b, and the N,N-di-n-propyl derivatives (+)-15b and (-)-15b demonstrate high affinity and selectivity for D<sub>1</sub> receptor. It is noteworthy that the enantiomers 14b bind streoselectively to  $D_1$  receptor; (+)-**14b** ( $K_i = 1.87$  nM) is 10 times more potent than (–)-**14b** ( $K_i = 18.68$  nM). The *N*,*N*-di-*n*-propyl derivatives show the same affinity for D<sub>1</sub> receptor, while the *N*-methyl-*N*-allyl and *N*-methyl-*N*-propyl derivatives (+)- and (-)-16b,17b are inactive at both receptor subtypes. It is of interest to note that in both series the N-alkylation with methyl and allyl or methyl and propyl groups greatly affects the  $D_1$ affinity; this increases in the 6-OH derivatives [(-)-16a], (+)-17a, and (-)-17a] and decreases in the 5-OH derivatives [ (+)-16b, (-)-16b, (+)-17b, and (-)-17b].

The structure of our compounds suggests that they can interact with the two primary binding sites of D<sub>1</sub> and D<sub>2</sub> receptors, namely an aspartate residue (amino group) and a serine residue (OH group). From the Dreiding model of 2-amino-5-hydroxy- and 2-amino-6hydroxy-1-phenyl-2,3-dihydro-1H-indene it appears that the distances between 5-OH or 6-OH and the nitrogen are quite similar. Therefore it is possible to hypothesize that the two hydroxyls bind at the same serine residue and, as a consequence, the arrangement of the unsubstituted phenyl ring could be different for the two series of compounds. Moreover, it seems that also the steric hindrance of the substituents on the amino group affects the binding. In the 6-OH indenes (+)- and (-)-14a-**17a**, showing  $D_1$  and  $D_2$  affinity, it may be hypothesized that the unsubstituted phenyl ring cannot properly fit the phenyl accessory binding site.<sup>8</sup> The size of the substituents influences the  $D_1$  affinity that increases when a propyl is replaced by a methyl group. On the other hand, for the 5-OH indenes the phenyl ring arrangement seems to be optimal for  $D_1$  interaction only in compounds (+)- and (-)-14b,15b, and D<sub>1</sub> affinity disappears when the N,N-di-n-propylamino group is replaced by a N-allyl-N-methylamino or N-methyl-N*n*-propylamino group. From this work it appears that the size of the substituents on the amino group greatly affects the affinity at DA receptors.

A previous study showed that the *trans*-2-amino-5,6dihydroxy-1-phenyl-2,3-dihydro-1*H*-indene (**18**, Table 1) does not displays affinity for DA receptors.<sup>23</sup> Since these results were not consistent with our data on 5-OH or 6-OH indenes, compound **18** as been resynthesized and tested with the same binding assay technique. Results obtained indicate that **18** binds selectively the D<sub>1</sub> receptor with a  $K_i$  of 5.5 nM and did not display D<sub>2</sub> receptor affinity. Binding experiments with this compound were made difficult to its extreme photosensitivity in aqueous solution. Hence, all procedures were done in a dark room. On the contrary our compounds, containing a phenol instead of a catechol group, are highly stable in solution.

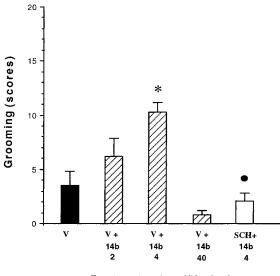
To further establish the pharmacological profile, attempting to gain information about their possible central agonist or antagonist activity, the compounds (+)-**14b** and (+)-**15b** (D<sub>1</sub> selective) were tested *in vivo* 



**Figure 1.** Effect of (*R*)-(+)-SCH 23390 on (+)-**15b** and (*R*)-(+)-SK&F 38393-induced grooming in rats. Vehicle [V, physiological (0.9%) saline], (+)-**15b**, and (*R*)-(+)-SK&F 38393 (SK&F) were intraperitoneally (ip) injected 5 min before the test (30 min); V and (*R*)-(+)-SCH 23390 (SCH) at 1.8  $\mu$ mol/kg were ip injected 25 min before V, (+)-**15b**, and SK&F. Histograms are the means ± SEM of the scores for grooming for each treatment group (at least eight animals). An asterisk (\*) shows results significantly different from those for vehicle-treated animals; a dot ( $\bullet$ ) indicates results significantly different from those of respective controls (Kruskal Wallis test followed by Mann Whitney *U* test).

with behavioral methods. It is well-known that  $D_1$ agonists induce prominent grooming in the rat, while pretreatment with D<sub>1</sub> antagonists reduces the grooming.<sup>24,25</sup> The intraperitoneal administration of (+)-15b  $(1.5-30 \,\mu \text{mol/kg})$  to rats significantly increased grooming, which was rather sporadic in vehicle-treated animals accustomed to the new cage (Figure 1). The effect was maximal at 3  $\mu$ mol/kg and disappeared completely at the dose of 30  $\mu$ mol/kg. Pretreatment with the DA D<sub>1</sub> antagonist (*R*)-(+)-SCH 23390 (1.8 µmol/kg) significantly counteracted the grooming response induced by 3  $\mu$ mol/kg of (+)-**15b**. The D<sub>1</sub> agonist (*R*)-(+)-7,8dihydroxy-1-phenyl-2,3,4,5-tetrahydro-1H-3-benzazepine (1b, SK&F 38393), comparatively tested at 3 and 30  $\mu$ mol/kg, enhanced grooming, and also in this case, the phenomenon was antagonized by (R)-(+)-SCH 23390 at 1.8  $\mu$ mol/kg. Figure 2 shows that also (+)-14b (2-40  $\mu$ mol/kg) significantly increased grooming at 4  $\mu$ mol/ kg with respect to vehicle, and the  $D_1$  antagonist (*R*)-(+)-SCH 23390 inhibited the grooming stimulation. However (+)-14b seems to be in vivo less active than (+)-15b. At all doses tested, (+)-14b, (+)-15b, and (*R*)-(+)-SK&F 38393 failed to induce other typical DA D<sub>2</sub> agonist responses<sup>26</sup> such as stereotyped behavior, stretching-yawning, and penile erection (data not shown). Thus, the behavioral response to (+)-14b and (+)-15b was induction of grooming that is characteristic of all preferential and selective  $D_1$  receptor agonists examined to date.<sup>24,27</sup> These preliminary studies also indicate that (+)-14b and (+)-15b cross blood-brain barrier and are active on central D<sub>1</sub> receptors.

In preliminary behavioral studies, the derivatives (-)-**15a** and (-)-**16a**, which displayed affinity for D<sub>1</sub> and D<sub>2</sub> receptors, did not promote stereotyped behavior, but induced some episodes of grooming not statistically



Treatments (µmol/Kg, i.p.)

**Figure 2.** Effect of (*R*)-(+)-SCH 23390 on (+)-**14b**-induced grooming in rats. Vehicle [V, physiological (0.9%) saline] and (+)-**14b** were intraperitoneally (ip) injected 5 min before the test (30 min); V and (*R*)-(+)-SCH 23390 (SCH) at 1.8  $\mu$ mol/kg were ip injected 25 min before V and (+)-**14b**. Histograms are the means ± SEM of the scores for grooming for each treatment group (at least eight animals). An asterisk (\*) denotes results significantly different from those for vehicle-treated animals; a dot ( $\bullet$ ) denotes results significantly different from those for respective controls (Kruskal Wallis test followed by Mann Whitney *U* test).

different with respect to vehicle. A similar effect was reported previously for compounds which showed affinity for  $D_1$  and  $D_2$  receptors.<sup>28</sup>

In summary, this report describes a series of trans-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indenes that in binding assays demonstrated affinity for both D<sub>1</sub> and  $D_2$  DA receptor subtypes. These data indicate that the putative pharmacophore of the  $D_{1-like}$  receptor,<sup>5</sup> the 2-phenyl-2-(3-hydroxyphenyl)ethylamine 5, embedded in these compounds in a trans extended conformation, does not induce high  $D_1$  selectivity. On the other hand, the isomeric trans-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1H-indenes and the di-N-n-propyl derivatives, containing the 2-phenyl-2-(4-hydroxyphenyl)ethylamine moiety, showed high affinity and selectivity for  $D_1$ receptor. In an "in vivo" behavioral study, (+)-14b and (+)-15b showed central D<sub>1</sub> agonistic activity. It is important to note that these compounds contain only a phenolic group, while most D<sub>1</sub> agonists known to date are catechol derivatives. These findings suggest that D<sub>1</sub> agonist activity does not require a catechol structure.

## **Experimental Section**

Melting points were determined on a Buchi 510 apparatus and are uncorrected. Microanalyses were performed on a 1106 Carlo Erba CHN analyzer, and the results were within  $\pm 0.4\%$  of the calculated values. <sup>1</sup>H NMR spectra were recorded on a Varian VXR 200 MHz spectrometer. Chemical shifts are reported in parts per million ( $\delta$ ) downfield from the internal standard tetramethylsilane (Me<sub>4</sub>Si). The IR spectra were run on a Perkin-Elmer Model 297 spectrometer as Nujol mulls or liquid films. The identity of all new compounds was confirmed by both elemental analysis and NMR data; homogeneity was confirmed by TLC on silica gel Merck 60 F<sub>254</sub>. Solutions were routinely dried over anhydrous sodium sulfate prior to evaporation. Chromatographic purifications were performed by Merck-60 silica gel columns 70–230 mesh ASTM from Merck

with a reported solvent. Optical rotations were obtained on a Perkin-Elmer 241 MC polarimeter.

(±)-trans-2-Amino-6-methoxy-1-phenyl-2,3-dihydro-1H-indene [(±)-6a] and (±)-trans-2-amino-5-methoxy-1phenyl-2,3-dihydro-1*H*-indene [(±)-6b]. Under nitrogen, 5(6)-methoxy-3-phenyl-1*H*-indene (5.55 g, 25 mmol) was added to a stirred solution of NaBH<sub>4</sub> (0.4 g, 10.3 mmol) in dry diglyme (15 mL). The flask was cooled in an ice bath, and boron trifluoride etherate (1.95 g, 13.75 mmol) was added dropwise. The solution was stirred at room temperature for 3 h. Then hydroxylamine O-sulfonic acid (3.2 g, 27.5 mmol) in dry diglyme (15 mL) was added, and the mixture was heated at 100 °C for 3 h. The solution was cooled, treated with concentrated HCl (10 mL), and poured into water (100 mL). The acidic solution was extracted with Et<sub>2</sub>O to remove diglyme and residue boronic acid, made basic with 2 N NaOH, and extracted with Et<sub>2</sub>O. The extracts were dried and evaporated. To the residue, dissolved in absolute EtOH, was added concentrated HCl (1.7 mL). Evaporation of the solvent gave a white solid which was recrystallized.  $(\pm)$ -**6a**·HCl: mp 236-238 °C (from *i*-PrOH), yield 33%. (±)-6b·HCl: mp 220-222 °C (from EtOH abs), yield 31%. The hydrochloride was treated with 15% NH<sub>4</sub>OH, and the amine was extracted with  $Et_2O$ . The extracts were dried and evaporated. The solid residue was recrystallized from Et<sub>2</sub>O.

(±)-6a: mp 87–88 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.28 (m, 5H, ArH), 6.76 (dd, 2H, H-5), 6.44 (d, 1H, H-7), 3.94 (d, 1H, J = 7.9 Hz, H-1), 3.70 (s, 3H, OCH<sub>3</sub>), 3.64 (q, 1H, H-2), 3.14 (dd, 1H, H-3), 2.76 (dd, 1H, H-3), 2.14 (s, 2H, NH<sub>2</sub>). Anal. (C<sub>16</sub>H<sub>17</sub>NO) C, H, N.

(±)-**6b**: mp 94–96 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.28 (m, 5H, ArH), 6.81 (d, 1H, H-7), 6.78 (s, 1H, H-4), 6.70 (m, 1H, H-6), 3.82 (d, 1H, J = 8 Hz, H-1), 3.80 (s, 3H, OCH<sub>3</sub>), 3.62 (q, 1H, H-2), 3.23 (dd, 1H, H-3), 2.85 (dd, 1H, H-3), 1.60 (s, 2H, NH<sub>2</sub>). Anal. (C<sub>16</sub>H<sub>17</sub>NO) C, H, N.

**Resolution of (±)-6a and (±)-6b.** A solution of (-)-2,3dibenzoyl-L-tartaric acid monohydrate (15.04 g, 40 mmol) in EtOH (60 mL) was added to a solution of (±)-**6a** (9.28 g, 39 mmol) in a mixture of EtOH (40 mL) and *i*-PrOH (60 mL). After standing at 0 °C for 24 h the solid was filtered and recrystallized from a mixture of EtOH (100 mL) and *i*-PrOH (40 mL). The colorless crystals (9.8 g) were recrystallized three times from a mixture of EtOH (100 mL) and *i*-PrOH (16 mL). (-)-2,3-Dibenzoyl-L-tartrate: mp 197–199 °C;  $[\alpha]^{20}_{D} = -74.8$ (*c* = 1, MeOH). Anal. (C<sub>16</sub>H<sub>17</sub>NO·C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>) C, H, N.

The crystalline salt was treated with an excess of 15% NH<sub>4</sub>-OH, and the solution was extracted with Et<sub>2</sub>O (large volume). After being dried the extracts were evaporated to give (+)-*trans*-2-amino-6-methoxy-1-phenyl-2,3-dihydro-1*H*-indene (+)-**6a**, which was rectrystallized from Et<sub>2</sub>O: mp 87–88 °C;  $[\alpha]^{20}_{D}$  = +33.67 (*c* = 1, EtOH); yield 2 g, 47%.

The ethanolic mother liquors from the preceding preparation of the (-)-2,3-dibenzoyl tartrate were concentrated *in vacuo*. A suspension of the residual solid in H<sub>2</sub>O was made alkaline with 15% NH<sub>4</sub>OH, and the mixture was extracted with Et<sub>2</sub>O. Concentration of the extracts gave the free base. To a solution of this free base (3.2 g, 13 mmol) in a mixture of EtOH (20 mL) and *i*-PrOH (40 mL) was added a solution of (+)-2,3dibenzoyl-L-tartaric acid (5.44 g, 14 mmol) in EtOH (40 mL). After 12 h at 0 °C the colorless crystals (5 g) were filtered and recrystallized three times from EtOH. (+)-2,3-Dibenzoyl-Ltartrate: mp 191–194 °C;  $[\alpha]^{20}_{\rm D}$  = +73.5 (*c* = 1, MeOH). Anal. (C<sub>16</sub>H<sub>17</sub>NO·C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>) C, H, N.

The crystalline salt was treated with an excess of 15% NH<sub>4</sub>-OH, and the solution was extracted with Et<sub>2</sub>O (large volume). After being dried the extracts were evaporated to give (–)-*trans*-2-amino-6-methoxy-1-phenyl-2,3-dihydro-1*H*-indene [(–)-**6a**], which was rectrystallized from Et<sub>2</sub>O: mp 87–88 °C;  $[\alpha]^{20}_{D}$  = -33.27 (*c* = 1, EtOH); yield 2.1 g, 51%.

A solution of (–)-2,3-dibenzoyl-L-tartaric acid monohydrate (15.04 g, 40 mmol) in EtOH (150 mL) was added to a solution of ( $\pm$ )-**6b** (9.28 g, 39 mmol) in EtOH (150 mL). After the mixture was left to stand at 0 °C for 24 h, the solid was filtered and recrystallized from a mixture of EtOH (190 mL) and MeOH (225 mL). The colorless crystals were recrystallized four times from EtOH. (–)-2,3-Dibenzoyl-L-tartrate: mp 195–

#### 2,3-Dihydro-1H-indenes as Dopamine Receptor Ligands

197 °C; [ $\alpha$ ]<sup>20</sup><sub>D</sub> = -85.6 (*c* = 1, MeOH); yield 2.7 g, 58%. Anal. (C<sub>16</sub>H<sub>17</sub>NO·C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>) C, H, N.

The crystalline salt was treated with an excess of 15% NH<sub>4</sub>-OH, and the solution was extracted with Et<sub>2</sub>O (large volume). After being dried the extracts were evaporated to give (+)-*trans*-2-amino-5-methoxy-1-phenyl-2,3-dihydro-1*H*-indene [(+)-**6b**], which was rectrystallized from Et<sub>2</sub>O: mp 93–96 °C; [ $\alpha$ ]<sup>20</sup><sub>D</sub> = +16.74 (*c* = 1, EtOH); yield 2.7 g, 58%.

The ethanolic mother liquors from the preceding preparation of the (–)-2,3-dibenzoyl tartrate were concentrated *in vacuo*. A suspension of the residual solid in H<sub>2</sub>O was made alkaline with 15% NH<sub>4</sub>OH, and the mixture was extracted with Et<sub>2</sub>O. Concentration of the extracts gave the free base. To a solution of this free base (5.80 g, 21 mmol) in MeOH (200 mL) was added a solution of (+)-2,3-dibenzoyl-L-tartaric acid (9.03 g, 25 mmol) in EtOH (200 mL). After 12 h at 0 °C the colorless crystals were filtered and recrystallized three times from EtOH/MeOH, 1/1. (+)-2,3-Dibenzoyl-L-tartrate: mp 195–196 °C;  $[\alpha]^{20}_{\rm D} = -83.6$  (c = 1, MeOH). Anal. (C<sub>16</sub>H<sub>17</sub>NO·C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>) C, H, N.

The crystalline salt was treated with an excess of 15% NH<sub>4</sub>-OH, and the solution was extracted with Et<sub>2</sub>O (large volume). After being dried, the extracts were evaporated to give (–)-*trans*-2-amino-5-methoxy-1-phenyl-2,3-dihydro-1*H*-indene [(–)-**6b**], which was recrystallized from Et<sub>2</sub>O: mp 93–96 °C;  $[\alpha]^{20}_{D} = -16.9$  (c = 1, EtOH); yield 2.8 g, 68%.

**Determination of the Enantiomeric Excess of the Amines (+)-6a, (-)-6a, (+)-6b, and (-)-6b.** Racemic and enantiomeric amines were derivatized with (*R*)-1-(1-naphthyl)ethyl isocyanate as described by Cannon.<sup>14</sup> Ureas were purified by column chromatography on silica gel with EtOAc/ *n*-hexane/*i*-PrOH (2/8/0.5) as eluent. NMR spectrum of racemic urea obtained from ( $\pm$ )-**6a** showed two doublet at 3.98 and 3.90 ppm (indene H-1 proton) and two doublet at 1.52 and 1.41 ppm (methyl protons). The NMR spectrum of the racemic urea obtained from ( $\pm$ )-**6b** showed two doublets at 3.94 and 3.85 ppm (indene H-1 proton) and two doublets at 1.52 and 1.42 ppm (methyl protons).

**1-(6(5)-Methoxy-1-phenyl-2,3-dihydro-1***H***-inden-2-yl)-3-(1-naphthalen-1-ylethyl)ureas (7a,b, 8a,b).** Inspection and integration of the benzylic and methyl protons signals in the NMR spectra provided an enantiomeric excess  $\geq$  98%.

**7a** was obtained from (+)-**6a**: mp 202–203 °C;  $[\alpha]^{20}_{\rm D} = -24.6$  (c = 1, EtOH); yield 88%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.11, 7.88 and 7.78 (three m, 3H, ArH), 7.52 and 7.38 (two m, 4H, ArH), 7.10 (m, 4H, ArH), 6.90 (m, 2H, ArH), 6.72 (dd, 1H, ArH), 6.38 (d, 1H, ArH), 5.55 (m, 1H, naphth-CH), 4.69 and 4.50 (two d, 2H, NH), 4.27 (m, 1H, H-2), 3.90 (d, 1H, H-1), 3.68 (s, 3H, OCH<sub>3</sub>), 3.34 and 2.61 (two dd, 2H, H-3), 1.52 (d, 3H, CH<sub>3</sub>). Anal. (C<sub>29</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>) C, H, N.

**8a** was obtained from (–)-**6a**: mp 190–191 °C;  $[\alpha]^{20}_{D} = -23.3$  (c = 1, EtOH); yield 89%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.06, 7.83 and 7.75 (three m, 3H, ArH), 7.49 (m, 2H, ArH), 7.32 (m, 5H, ArH), 7.13 (m, 3H, ArH), 6.72 (dd, 1H, ArH), 6.40 (d, 1H, ArH), 5.53 (m, 1H, naphth-CH), 4.66 and 4.43 (two d, 2H, NH), 4.25 (m, 1H, H-2), 3.98 (d, 1H, H-1), 3.68 (s, 3H, OCH<sub>3</sub>), 3.28 and 2.56 (two dd, 2H, H-3), 1.41 (d, 3H, CH<sub>3</sub>). Anal. (C<sub>29</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>) C, H, N.

**7b** was obtained from (+)-**6b**: mp 218–219 °C;  $[\alpha]^{20}_{D} = -22.5$  (c = 1, EtOH); yield 88%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.10, 7.88 and 7.78 (three m, 3H, ArH), 7.51 and 7.38 (two m, 4H, ArH), 7.08 (m, 3H, ArH), 6.89 (m, 2H, ArH), 6.70 (m, 3H, ArH), 5.56 (m, 1H, naphth-CH), 4.52 (bs, 2H, NH), 4.24 (m, 1H, H-2), 3.85 (d, 1H, H-1), 3.78 (s, 3H, OCH<sub>3</sub>), 3.37 and 2.65 (two dd, 2H, H-3), 1.52 (d, 3H, CH<sub>3</sub>). Anal. ( $C_{29}H_{28}N_2O_5$ ) C, H, N.

**8b** was obtained from (–)-**6b**: mp 203–204 °C;  $[\alpha]^{20}_{D} = +10$  (c = 1, EtOH); yield 80%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.05, 7.83, and 7.74 (three m, 3H, ArH), 7.48 and 7.38 (two m, 4H, ArH), 7.28 (m, 3H, ArH), 7.12 (m, 2H, ArH), 6.71 (m, 3H, ArH), 5.53 (m, 1H, naphth-CH), 4.62 and 4.41 (two bs, 2H, NH), 4.23 (m, 1H, H-2), 3.94 (d, 1H, H-1), 3.78 (s, 3H, OCH<sub>3</sub>), 3.32 and 2.61 (two dd, 2H, H-3), 1.42 (d, 3H, CH<sub>3</sub>). Anal. (C<sub>29</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>) C, H, N.

*trans*-6-Methoxy-1-phenyl-2-(di-*n*-propylamino)-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-9a, (-)-9a] and *trans*-5-methoxy-1-phenyl-2-(di-*n*-propylamino)-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-9b, (-)-9b]. The N- alkylation of amines (+)-**6a**, (-)-**6a**, (+)-**6b**, and (-)-**6b** was accomplished as described by Marchini et al.<sup>15</sup> The amine (2 g, 8.4 mmol) was allowed to react with sodium borohydride (3.15 g, 84 mmol) and propionic acid (10.4 g, 140 mmol) in dry benzene (40 mL). The oily residue was chromatographed on silica gel column eluted with EtOAc/cyclohexane, 75/25 [(+)-**9a**, (-)-**9a**] or EtOAc [(+)-**9b**, (-)-**9b**]. The hydrochlorides were prepared by addition of 37% HCl to a solution of the base in EtOH. The solvent was evaporated under reduced pressure, and the residue was recrystallized from EtOAc/ Et<sub>2</sub>O.

(+)-**9a·HCl** was obtained from (-)-**6a**: mp 148–151 °C; [ $\alpha$ ]<sup>20</sup><sub>D</sub> = +64.35 (*c* = 1, EtOH); yield 50%; <sup>1</sup>H NMR (DMSO *d*<sub>6</sub>)  $\delta$  10.74 (bs, 1H, NH), 7.40 (m, 5H, ArH), 7.27 (d, 1H, H-4), 6.85 (dd, 1H, H-5), 6.15 (d, 1H, H-7), 4.86 (d, 1H, H-1), 4.50 (m, 1H, H-2), 3.62 (s, 3H, OCH<sub>3</sub>), 3.51 and 3.32 (two m, 2H, H-3), 3.0 (m, 4H, NCH<sub>2</sub>C), 1.68 and 1.37 (two m, 4H, NCCH<sub>2</sub>), 0.89 and 0.53 (two t, 6H, CH<sub>3</sub>). Anal. (C<sub>22</sub>H<sub>30</sub>ClNO) C, H, N.

(-)-**9a**·HCl was obtained from (+)-**6a**: mp 148–151 °C;  $[\alpha]^{20}_D$  = -64.03 (c = 1, EtOH); yield 55%. The NMR spectrum was the same as that of (+)-**9a**·HCl. Anal. (C<sub>22</sub>H<sub>30</sub>ClNO) C, H, N.

(+)-**9b**·HCl was obtained from (-)-**6b**: mp 159–160 °C; [ $\alpha$ ]<sup>20</sup><sub>D</sub> = +88.98 (*c* = 1, EtOH); yield 50%; <sup>1</sup>H NMR (DMSO *d*<sub>6</sub>)  $\delta$  11.0 (bs, 1H, NH), 7.37 (m, 5H, ArH), 6.90 (d, 1H, H-4), 6.72 (dd, 1H, H-6), 6.53 (d, 1H, H-7), 4.81 (d, 1H, H-1), 4.49 (m, 1H, H-2), 3.75 (s, 3H, OCH<sub>3</sub>), 3.56 and 3.39 (two m, 2H, H-3), 2.98 (m, 4H, NCH<sub>2</sub>C), 1.68 and 1.36 (two m, 4H, NCCH<sub>2</sub>), 0.88 and 0.53 (two t, 6H, CH<sub>3</sub>). Anal. (C<sub>22</sub>H<sub>30</sub>ClNO) C, H, N.

(-)-**9b**·HCl was obtained from (+)-**6b**: mp 159–160 °C;  $[\alpha]^{20}{}_{D} = -89.5$  (c = 1, EtOH); yield 45%. The NMR spectrum was the same as that of (+)-**9b**·HCl. Anal. (C<sub>22</sub>H<sub>30</sub>ClNO) C, H, N.

*trans*-6-Methoxy-1-phenyl-2-[(ethoxycarbonyl)amino]-2,3-dihydro-1*H*-indenes [(+)-10a, (-)-10a] and *trans*-5methoxy-1-phenyl-2-[(ethoxycarbonyl)amino]-2,3-dihydro-1*H*-indenes [(+)-10b, (-)-10b]. A solution of ethyl chloroformate (1.18 g, 11 mmol) in anhydrous Et<sub>2</sub>O (15 mL) was added dropwise to a solution of the amines (+)-6a, (-)-6a, (+)-6b, and (-)-6b (2.6 g, 11 mmol) in anhydrous Et<sub>2</sub>O (70 mL) and triethylamine (3.1 mL, 22 mmol), cooled at 0 °C. The reaction mixture was allowed to reach room temperature and then stirred for 1 h. Water was then added, and the aqueous solution was extracted with three portions of CHCl<sub>3</sub>. The combined organic phases were dried and evaporated. The solid residue was recrystallized.

(+)-**10a** was obtained from (+)-**6a**: mp 95–96 °C (from cyclohexane);  $[\alpha]^{20}{}_{\rm D}$  = +15.4 (*c* = 1, EtOH); yield 92%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.27 (m, 6H, ArH), 6.80 (dd, 1H, H-5), 6.53 (d, 1H, H-7), 4.94 (bs, 1H, NH), 4.34 (m, 1H, H-2), 4.10 (m, 3H, H-1, OCH<sub>2</sub>), 3.72 (s, 3H, OCH<sub>3</sub>), 3.36 (dd, 1H, H-3), 2.74 (dd, 1H, H-3), 1.20 (t, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>21</sub>NO<sub>3</sub>) C, H, N.

(–)-10a was obtained from (–)-6a: mp 95–96 °C (from cyclohexane);  $[\alpha]^{20}{}_D=-14.95$  (c = 1, EtOH); yield 87%. The NMR spectrum was the same as that of (+)-10a. Anal. (C<sub>19</sub>H<sub>21</sub>NO<sub>3</sub>) C, H, N.

(+)-10b was obtained from (–)-6b: mp 174–176 °C (from *i*-PrOH);  $[\alpha]^{20}{}_{\rm D}$  = +16.1 (*c* = 1, EtOH); yield 81%; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.23 (m, 5H, ArH), 6.85 (m, 2H, ArH), 6.75 (dd, 1H, H-6), 4.94 (bs, 1H, NH), 4.35 (m, 1H, H-2), 4.09 (m, 3H, H-1, OCH<sub>2</sub>), 3.82 (s, 3H, OCH<sub>3</sub>), 3.40 (dd, 1H, H-3), 2.80 (dd, 1H, H-3), 1.22 (t, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>21</sub>NO<sub>3</sub>) C, H, N.

(-)-10b was obtained from (+)-6b: mp 174–176 °C (from i-PrOH);  $[\alpha]^{20}_D = -15.62$  (c = 1, EtOH); yield 81%. The NMR spectrum was the same as that of (+)-10b. Anal. ( $C_{19}H_{21}$ -NO<sub>3</sub>) C, H, N.

*trans***6**-Methoxy-2-(methylamino)-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-11a, (-)-11a] and *trans*-5-methoxy-2-(methylamino)-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-11b, (-)-11b]. A solution of the compounds (+)-10a, (-)-10a, (+)-10b, and (-)-10b (3 g, 9.6 mmol) in anhydrous THF (40 mL) was added dropwise to a stirred suspension of LiAlH<sub>4</sub> (0.7 g, 18.4 mmol) in anhydrous Et<sub>2</sub>O (70 mL), under a nitrogen atmosphere. The mixture was heated to 40 °C for 24 h. The reaction was then terminated by the addition of water (0.7 mL), 15% NaOH (0.7 mL), and finally water (2.1 mL). The solid was filtered and washed with  $Et_2O$ . The filtrates were dried and evaporated. To the oily residue, dissolved in EtOH (60 mL), was added 37% HCl (1.6 mL). The solid obtained after evaporation of the solvent was recrystallized.

(+)-**11a**·HCl was obtained from (–)-**10a**: mp 182–184 °C (from *i*·PrOH/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +26.3$  (c = 1, EtOH); yield 73%; <sup>1</sup>H NMR (DMSO- $d_{6}$ )  $\delta$  9.52 and 9.32 (two bs, 2H, NH<sub>2</sub><sup>+</sup>), 7.33 (m, 6H, ArH), 6.86 (dd, 1H, H-5), 6.36 (d, 1H, H-7), 4.66 (d, 1H, H-1), 4.0 (m, 1H, H-2), 3.65 (s, 3H, OCH<sub>3</sub>), 3.30 (dd, 1H, H-3), 3.10 (dd, 1H, H-3), 2.53 (s, 3H, CH<sub>3</sub>). Anal. (C<sub>17</sub>H<sub>20</sub>-ClNO) C, H, N.

(-)-**11a**·HCl was obtained from (+)-**10a**: mp 182–184 °C (from *i*·PrOH/Et<sub>2</sub>O); [ $\alpha$ ]<sup>20</sup><sub>D</sub> = -26.10 (*c* = 1, EtOH); yield 72%. The NMR spectrum was the same as that of (+)-**11a**·HCl. Anal. (C<sub>17</sub>H<sub>20</sub>ClNO) C, H, N.

(+)-11b·HCl was obtained from (+)-10b: mp 239–241 °C (from *i*-PrOH);  $[\alpha]^{20}{}_{D} = +52.16$  (c = 1, EtOH); yield 75%. <sup>1</sup>H NMR (DMSO-d<sub>6</sub>)  $\delta$  9.59 and 9.40 (two bs, 2H, NH<sub>2</sub><sup>+</sup>), 7.33 (m, 5H, ArH), 6.95 (d, 1H, H-4), 6.75 (m, 2H, ArH), 4.62 (d, 1H, H-1), 4.01 (m, 1H, H-2), 3.76 (s, 3H, OCH<sub>3</sub>), 3.45 (dd, 1H, H-3), 3.18 (dd, 1H, H-3), 2.52 (s, 3H, CH<sub>3</sub>). Anal. (C<sub>17</sub>H<sub>20</sub>ClNO) C, H, N.

(-)-**11b**·HCl was obtained from (-)-**10b**: mp 239–241 °C (from *i*-PrOH);  $[\alpha]^{20}_D = -51.79$  (c = 1, EtOH); yield 73%. The NMR spectrum was the same as that of (+)-**11b**·HCl. Anal. (C<sub>17</sub>H<sub>20</sub>ClNO) C, H, N.

trans-N-Methyl-N-allyl-2-amino-6-methoxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-12a, (-)-12a] and trans-N-methyl-N-allyl-2-amino-5-methoxy-1-phenyl-2,3-dihydro-1H-indene Hydrochlorides [(+)-12b, (-)-12b]. A mixture of the hydrochlorides (+)-11a, (-)-11a, (+)-11b, and (-)-11b (1.5 g, 5.2 mmol) in absolute EtOH (35 mL), anhydrous K<sub>2</sub>CO<sub>3</sub> (0.77 g, 5.6 mmol), and allyl bromide (1.22 g, 10 mmol) was stirred at 60 °C for 3 h. After removal of the solvent in vacuum, H<sub>2</sub>O was added, and the mixture was extracted with Et<sub>2</sub>O. The combined organic extracts were dried, filtered, and evaporated. The oily residue was purified by column chromatography with EtOAc as eluent. The desired fraction was collected and evaporated, the residue was dissolved in EtOH (35 mL), and 37% HCl (1.3 mL) was added. The solid obtained after evaporation of the solvent was recrystallized from EtOAc/Et<sub>2</sub>O.

(+)-**12a**·HCl was obtained from (+)-**11a**·HCl: mp 198–200 °C;  $[\alpha]^{20}_{D} = +57.0 \ (c = 1, EtOH)$ ; yield 65%. <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  11.67 (bs, 1H, NH<sup>+</sup>), 7.33 (m, 6H, ArH), 6.84 (d, 1H, H-7), 6.30 (dd, 1H, H-5), 5.92 (m, 1H, CH=), 5.40 (m, 2H, =CH<sub>2</sub>), 4.91 (d, 1H, H-1), 4.28 (m, 1H, H-2), 3.77 (m, 1H, H-3), 3.61 (s, 3H, OCH<sub>3</sub>), 3.46 (m, 3H, H-3, NCH<sub>2</sub>), 2.60 and 2.52 (two d, 3H, CH<sub>3</sub>). Anal. (C<sub>20</sub>H<sub>24</sub>ClNO) C, H, N.

(-)-**12a**·HCl was obtained from (-)-**11a**·HCl: mp 198–200 °C;  $[\alpha]^{20}_{D} = +57.30$  (c = 1, EtOH); yield 64%.The NMR spectrum was the same as that of (+)-**12a**·HCl. Anal. (C<sub>20</sub>H<sub>24</sub>-ClNO) C, H, N.

(+)-**12b**·HCl was obtained from (+)-**11b**·HCl: mp 68–70 °C; [α]<sup>20</sup><sub>D</sub> = +80.22 (*c* = 1, EtOH); yield 74%. <sup>1</sup>H NMR (DMSO *d*<sub>6</sub>) δ 11.53 (bs, 1H, NH<sup>+</sup>), 7.34 (m, 5H, ArH), 6.93 (d, 1H, H-4), 6.71 (m, 2H, ArH), 5.93 (m, 1H, CH=), 5.40 (m, 2H, =CH<sub>2</sub>), 4.82 (d, 1H, H-1), 4.28 (m, 1H, H-2), 3.80 (m, 1H, H-3), 3.72 (s, 3H, OCH<sub>3</sub>), 3.49 (m, 3H, H-3, NCH<sub>2</sub>), 3.37 (m, 1H, H-3), 2.62 and 2.53 (two d, 3H, CH<sub>3</sub>). Anal. (C<sub>20</sub>H<sub>24</sub>ClNO) C, H, N. (-)-**12b**·HCl was obtained from (-)-**11b**·HCl: mp 68–70 °C;

 $[\alpha]^{20}_{D} = -79.90 \ (c = 1, EtOH); yield 70\%. The NMR spectrum was the same as (+)-12b·HCl. Anal. (C<sub>20</sub>H<sub>24</sub>ClNO) C, H, N.$ 

*trans*-*N*-Methyl-*N*-*n*-propyl-2-amino-6-methoxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-13a, (-)-13a] and *trans*-*N*-methyl-*N*-propyl-2-amino-5-methoxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-13b, (-)-13b]. A mixture of the hydrochlorides (+)-11a, (-)-11a, (+)-11b, and (-)-11b (1.5 g, 5.2 mmol) in acetone (80 mL), anhydrous K<sub>2</sub>CO<sub>3</sub> (2.15 g, 15.5 mmol), and iodopropane (1.77 g, 10.4 mmol) was stirred to reflux for 3 h. At the end of this period another portion of iodopropane (0.9 g, 10 mmol) was added, and the suspension was refluxed for 2 h. After removal of solvent in vacuum, water was added, and the mixture was extracted with CHCl<sub>3</sub>. The combined organic extracts were dried, filtered, and evaporated. The oily residue was purified by column chromatography with EtOAc as eluent. The desired fraction was collected and evaporated. The residue was dissolved in EtOH (35 mL), and 37% HCl (1.3 mL) was added. The solid obtained after evaporation of the solvent was recrystallized from EtOAc/Et<sub>2</sub>O.

(+)-**13a**·HCl was obtained from (+)-**11a**·HCl: mp 175–176 °C;  $[\alpha]^{20}_{D} = +60.10$  (*c* = 1, EtOH); yield 86%; <sup>1</sup>H NMR (DMSO*d*<sub>6</sub>)  $\delta$  11.17 and 11.01 (two bs, 1H, NH<sup>+</sup>), 7.33 (m, 6H, ArH), 6.84 (dd, 1H, H-5), 6.28 (d, 1H, H-7), 4.85 (m, 1H, H-1), 4.34 (m, 1H, H-2), 3.62 (s, 3H, OCH<sub>3</sub>), 3.41 (m, 2H, H-3), 2.98 (m, 2H, NCH<sub>2</sub>), 2.69 and 2.60 (two d, 3H, NCH<sub>3</sub>), 1.65 and 1.43 (two m, 2H, CCH<sub>2</sub>C), 0.88 and 0.64 (two t, 3H, CH<sub>3</sub>). Anal. (C<sub>20</sub>H<sub>26</sub>CINO) C, H, N.

(-)-**13a**·HCl was obtained from (-)-**11a**·HCl: mp 175–176 °C;  $[\alpha]^{20}_D = -60.40$  (c = 1, EtOH); yield 90%. The NMR spectrum was the same as that of (+)-**13a**·HCl. Anal. (C<sub>20</sub>H<sub>26</sub>-ClNO) C, H, N.

(+)-13b·HCl was obtained from (+)-11b·HCl: mp 150–152 °C;  $[\alpha]^{20}{}_{\rm D}$  = +78.30 (*c* = 1, EtOH); yield 89%. <sup>1</sup>H NMR (DMSO*d*<sub>6</sub>)  $\delta$  11.17 and 11.0 (two bs, 1H, NH<sup>+</sup>), 7.38 (m, 5H, ArH), 6.96 (d, 1H, H-4), 6.72 (m, 2H, H-6,7), 4.81 (d, 1H, H-1), 4.38 (m, 1H, H-2), 3.76 (s, 3H, OCH<sub>3</sub>), 3.44 (m, 2H, H-3), 2.98 (m, 2H, NCH<sub>2</sub>), 2.72 and 2.64 (two d, 3H, NCH<sub>3</sub>), 1.69 and 1.42 (two m, 2H, CCH<sub>2</sub>C), 0.90 and 0.68 (two t, 3H, CH<sub>3</sub>). Anal. (C<sub>20</sub>H<sub>26</sub>ClNO) C, H, N.

(-)-**13b**·HCl was obtained from (-)-**11b**·HCl: mp 150–152 °C;  $[\alpha]^{20}_{D} = -77.60$  (*c* = 1, EtOH); yield 86%. The NMR spectrum was the same as that of (+)-**13b**·HCl. Anal. (C<sub>20</sub>H<sub>26</sub>-ClNO) C, H, N.

**General Procedure for the Demethylation.** Methanesulfonic acid (71 mL) and methionine (9.8 g, 65.7 mmol) were added to a solution of the metoxyamine hydrochloride (1.6 g, 5.8 mmol) in water (4.5 mL). The mixture was stirred at 25 °C for 4 days, then poured into ice-water (80 mL), and made basic (pH = 8) with 15% NH<sub>4</sub>OH. The mixture was extracted with EtOAc. The organic extracts, washed with aqueous NaHSO<sub>3</sub> and water, were dried and evaporated. The residue was dissolved in EtOH (20 mL), and 37% HCl (0.9 mL) was added. The solid obtained after evaporation of the solvent was recrystallized.

*trans*-2-Amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-14a and (-)-14a]. (+)-14a·HCl was obtained from (-)-6a·HCl: mp 249–251 °C (from EtOAc/ Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +5.30$  (c = 1, EtOH); yield 70%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  9.30 (bs, 1H, OH), 8.49 (bs, 3H, NH<sub>3</sub><sup>+</sup>), 7.30 (m, 5H, ArH), 7.13 (d, 1H, H-4), 6.68 (dd, 1H, H-5), 6.23 (d, 1H, H-7), 4.44 (d, 1H, H-1), 3.88 (m, 1H, H-2), 3.32 (dd, 1H, H-3), 2.97 (dd, 1H, H-3). Anal. (C<sub>15</sub>H<sub>16</sub>ClNO) C, H, N.

(-)-**14a**·HCl was obtained from (+)-**6a**·HCl: mp 249–251 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = -5.21$  (c = 1, EtOH); yield 66%. The NMR spectrum was the same as that of (+)-**14a**·HCl. Anal. (C<sub>15</sub>H<sub>16</sub>ClNO) C, H, N.

*trans*-2-Amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-14b and (-)-14b]. (+)-14b·HCl was obtained from (-)-6b·HCl: mp 245-247 °C (from *i*-PrOH/ Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +17.72$  (c = 1, EtOH); yield 44%; <sup>1</sup>H NMR (DMSO- $d_{6}$ )  $\delta$  9.46 (s, 1H, OH), 8.46 (bs, 3H, NH<sub>3</sub><sup>+</sup>), 7.33 (m, 3H, ArH), 7.18 (m, 2H, ArH), 6.73 (d, 1H, H-4), 6.62 (m, 2H, H-6,7), 4.37 (d, 1H, H-1), 3.88 (m, 1H, H-2), 3.35 (m, 1H, H-3), 2.94 (m, 1H, H-3). Anal. (C<sub>15</sub>H<sub>16</sub>ClNO) C, H, N.

(-)-**14b**·HCl was obtained from (+)-**6b**·HCl: mp 245–247 °C (from *i*·PrOH/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = -17.60$  (c = 1, EtOH); yield 48%. The NMR spectrum was the same as that of (+)-**14b**·HCl. Anal. (C<sub>15</sub>H<sub>16</sub>ClNO) C, H, N.

*trans-N,N*-Di-*n*-propyl-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-15a, (-)-15a]. (+)-15a·HCl was obtained from (+)-9a·HCl: mp 226–228 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +70.0$  (c = 1, EtOH); yield 86%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  10.80 (bs, 1H, NH<sup>+</sup>), 9.26 (s, 1H, OH), 7.40 (m, 5H, ArH), 7.10 (d, 1H, H-4), 6.66 (dd, 1H, H-5), 6.02 (d, 1H, H-7), 4.80 (d, 1H, H-1), 4.45 (m, 1H, H-2), 3.45 (m, 1H, H-3), 3.24 (m, 1H, H-3), 2.92 (m, 4H, N-CH<sub>2</sub>), 1.65 and 1.32 (two m, 4H, NCCH<sub>2</sub>), 0.88 and 0.52 (two t, 6H, CH<sub>3</sub>). Anal. (C<sub>21</sub>H<sub>28</sub>CINO) C, H, N.

(–)-**15a**·HCl was obtained from (–)-**9a**·HCl: mp 226–228 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = -69.60$  (c = 1, EtOH); yield

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58%. The NMR spectrum was the same as (+)-**15a**·HCl. Anal. ( $C_{21}H_{28}CINO$ ) C, H, N.

*trans-N,N*-Di-*n*-propyl-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-15b, (-)-15b]. (+)-15b-HCl was obtained from (+)-9b-HCl: mp 237– 239 °C (from *i*-PrOH);  $[\alpha]^{20}_{D} = +86.87$  (c = 1, EtOH); yield 58%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  11.16 (bs, 1H, NH<sup>+</sup>), 9.48 (s, 1H, OH), 7.40 (m, 5H, ArH), 6.72 (d, 1H, H-4), 6.58 (dd, 1H, H-6), 6.40 (d, 1H, H-7), 4.78 (d, 1H, H-1), 4.40 (m, 1H, H-2), 3.41 (m, 2H, H-3), 2.94 (m, 4H, N-CH<sub>2</sub>), 1.65 and 1.32 (two m, 4H, NCCH<sub>2</sub>), 0.90 and 0.52 (two t, 6H, CH<sub>3</sub>). Anal. (C<sub>21</sub>H<sub>28</sub>CINO) C, H, N.

(-)-**15b**·HCl was obtained from (-)-**9b**·HCl: mp 238–239 °C (from *i*·PrOH);  $[\alpha]^{20}_{\rm D} = -87.21$  (*c* = 1, EtOH); yield 58%. The NMR spectrum was the same as (+)-**15b**·HCl. Anal. (C<sub>21</sub>H<sub>28</sub>ClNO) C, H, N.

*trans*-*N*-Methyl-*N*-allyl-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-16a, (-)-16a]. (+)-16a·HCl was obtained from (+)-12a·HCl: mp 107–109 °C (from EtOH/Et<sub>2</sub>O, hygroscopic);  $[\alpha]^{20}_{D} = +64.20$  (c = 1, EtOH); yield 71%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  11.18 (bs, 1H, NH<sup>+</sup>), 9.30 (s, 1H, OH), 7.35 (m, 5H, ArH), 7.13 (d, 1H, H-4), 6.67 (dd, 1H, H-7), 6.18 (m, 1H, H-5), 5.88 (m, 1H, CH=), 5.41 (m, 2H, CH<sub>2</sub>=), 4.81 (d, 1H, H-1), 4.30 (m, 1H, H-2), 3.68 (m, 1H, H-3), 3.48 (m, 3H, H-3, N-CH<sub>2</sub>), 2.69 and 2.53 (two d, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>22</sub>ClNO) C, H, N.

(-)-**16a**·HCl was obtained from (-)-**12a**·HCl: mp 107–109 °C (from EtOH/Et<sub>2</sub>O, hygroscopic);  $[\alpha]^{20}_{D} = -64.30$  (c = 1, EtOH); yield 72%. The NMR spectrum was the same as that of (+)-**16a**·HCl. Anal. (C<sub>19</sub>H<sub>22</sub>ClNO) C, H, N.

*trans*-*N*-Methyl-*N*-allyl-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-16b, (–)-16b]. (+)-16b·HCl was obtained from (+)-12b·HCl: mp 87– 89 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +75.93$  (c = 1, EtOH); yield 74%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  11.39 (bs, 1H, NH<sup>+</sup>), 9.50 (s, 1H, OH), 7.32 (m, 5H, ArH), 6.71 (d, 1H, H-4), 6.58 (m, 2H, H-6,7), 5.89 (m, 1H, CH=), 5.42 (m, 2H, CH<sub>2</sub>=), 4.78 (d, 1H, H-1), 4.22 (m, 1H, H-2), 3.77 (m, 2H, H-3), 3.40 (m, 2H, N-CH<sub>2</sub>), 2.63 and 2.53 (two d, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>22</sub>ClNO) C, H, N.

(-)-**16b**·HCl was obtained from (-)-**12b**·HCl: mp 86–88 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = -76.70$  (c = 1, EtOH); yield 84%. The NMR spectrum was the same as that of (+)-**16b**·HCl. Anal. (C<sub>19</sub>H<sub>22</sub>ClNO) C, H, N.

*trans*-*N*-Methyl-*N*-*n*-propyl-2-amino-6-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-17a, (-)-17a]. (+)-17a·HCl was obtained from (+)-13a·HCl: mp 117– 119 °C (from EtOH/Et<sub>2</sub>O, hygroscopic);  $[\alpha]^{20}{}_{\rm D}$  = +64.40 (c = 1, EtOH); yield 69%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  10.83 and 10.71 (two bs, 1H, NH<sup>+</sup>), 9.30 (s, 1H, OH), 7.34 (m, 5H, ArH), 7.14 (d, 1H, H-4), 6.67 (dd, 1H, H-5), 6.12 (m, 1H, H-7), 4.76 (d, 1H, H-1), 4.36 (m, 1H, H-2), 3.33 (m, 2H, H-3), 3.0 (m, 2H, N-CH<sub>2</sub>), 2.75 and 2.60 (two d, 3H, NCH<sub>3</sub>), 1.65 and 1.41 (two m, 2H, CCH<sub>2</sub>C), 0.90 and 0.66 (two t, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>24</sub>-CINO) C, H, N.

(-)-**17a**·HCl was obtained from (-)-**13a**·HCl: mp 117–119 °C (from EtOH/Et<sub>2</sub>O, hygroscopic);  $[\alpha]^{20}_D = -64.60$  (c = 1, EtOH); yield 73%. The NMR spectrum was the same as that of (+)-**17a**·HCl. Anal. (C<sub>19</sub>H<sub>24</sub>ClNO) C, H, N.

*trans*-*N*-Methyl-*N*-propyl-2-amino-5-hydroxy-1-phenyl-2,3-dihydro-1*H*-indene Hydrochlorides [(+)-17b, (-)-17b]. (+)-17b·HCl was obtained from (+)-13b·HCl: mp 128– 130 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = +75.50$  (c = 1, EtOH); yield 74%; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  11.18 and 11.04 (two bs, 1H, NH<sup>+</sup>), 9.47 (s, 1H, OH), 7.31 (m, 5H, ArH), 6.72 (d, 1H, H-4), 6.55 (m, 2H, H-6,7), 4.72 (m, 1H, H-1), 4.29 (m, 1H, H-2), 3.40 (m, 2H, H-3), 2.98 (m, 2H, N-CH<sub>2</sub>), 2.69 and 2.59 (two d, 3H, NCH<sub>3</sub>), 1.67 and 1.43 (two m, 2H, CCH<sub>2</sub>C), 0.88 and 0.63 (two t, 3H, CH<sub>3</sub>). Anal. (C<sub>19</sub>H<sub>24</sub>CINO) C, H, N.

(-)-17**b**·HCl was obtained from (-)-13**b**·HCl: mp 128–130 °C (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{20}_{D} = -76.40$  (c = 1, EtOH); yield 76%. The NMR spectrum was the same as that of (+)-17**b**·HCl. Anal. (C<sub>19</sub>H<sub>22</sub>ClNO) C, H, N.

**Pharmacology. Materials.** (R)-(+)-SCH 23390 hydrochloride, ( $\pm$ )-PPHT hydrochloride, and (R)-(+)-SK&F 38393 hydrochloride were purchased from Research Biochemicals International (Natick, MA). Dopamine was purchased from Sigma Chemical Co. (St. Louis, MO). [<sup>3</sup>H]Spiperone and [<sup>3</sup>H]-SCH 23390 were purchased from Amersham Radiochemical Centre (Buckinghamshire, UK). All substances employed in the binding assays were dissolved in distilled water. For behavioral tests the compounds were dissolved in physiological (0.9%) saline and injected intraperitoneally (ip).

**Animals.** In the radioligand-binding studies striata were obtained from male Sprague–Dawley rats (250–300 g body weight) obtained from Charles River (Calco, Italy). Behavioral studies were performed on male Wistar rats (Morini, S. Polo d'Enza, Italy) weighing 250–280 g.

**Receptor Binding.** The compounds were evaluated for their *in vitro* binding affinity at the  $D_1$  receptor with [<sup>3</sup>H]SCH 23390 (70 Ci/mmol, 0.25 nM) as a ligand, and at the  $D_2$  receptor using [<sup>3</sup>H]spiperone (42 Ci/mmol, 0.25 nM) as a ligand. Binding experiments were performed according to the protocol detailed in a former study of our group.<sup>17</sup>

**Behavioral Studies.** Animals were housed in groups of four with food and water available ad libitum, and a 24 h light cycle, from 7 a.m. to 7 p.m., for at least 1 week prior to the start of the tests. All the behavioral procedures were performed between 9 a.m. and 1 p.m. in a soundproof, airconditioned room, the animals being monitored by trained observers unaware of the experimental design. Twenty minutes before the start of the experiments, the animals were transferred, in groups of four, to glass observation cages (40 × 30 × 34 cm) to accustom them to the new environment. The test started immediately after the last treatment and lasted 30 min.

The experiments were replicated on separate days using new groups of animals. Grooming was evaluated according to Gispen et al.<sup>29</sup> In brief, an observer recorded every 15 s whether or not each rat displayed the phenomenon defined as follows: face and body washing, scratching, licking paws or tail. If one of these signs was observed, a positive score was given. In the case of antagonism experiments, the dose used for SCH 23390 was chosen on the basis of previous works<sup>24</sup> and was "per se" ineffective to modify the behaviour in question with respect to saline injection. Data for grooming are presented as mean  $\pm$  SEM of the cumulative scores for each animal during the test period and were analyzed using Kruskal Wallis test followed by Mann Whitney *U* test, with the level of significance set at p < 0.05.

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